

# M5 Protein A/G Agarose 使用说明书

产品名称	单位	货号	
M5 Protein A/G Agarose	1ml	MF099-01	

## [STORAGE]

Store at 4°C. Stable for one year from the date of shipment.

### [BACKGROUND]

The Protein A/G-Agarose is provided as an agarose conjugate for use in immunoprecipitation or purification. The product is provided as 0.5 ml agarose in 1 ml PBS buffer with 0.05% Sodium Azide. Protein A/G-Agarose is preblocked with BSA to reduce non-specific immunoglobulin binding. Sufficient product is provided for 25-33 immunoprecipitation reactions, to be used at 30-40  $\mu$ l resuspended volume per reaction. Protein A/G is a genetically engineered protein (MW ~ 50,500; apparent MW by SDS-PAGE ~ 40,000-45,000) that combines the IgG binding profiles of both Protein A and Protein G. The secreted Protein A/G contains four Fc-binding domains from Protein A and two from Protein G. In addition, Protein A/G binding to immunoglobulins is not as pH dependent as Protein A.

### [DESCRIPTION]

Protein A/G-Agarose is suitable for immunoprecipitation of mouse IgG1, IgG2a, IgG2b and IgG3, rat IgG1, IgG2a, IgG2b and IgG2c, rabbit and goat polyclonal Abs, and human IgG1, IgG2, IgG3 and IgG4.

#### [APPLICATIONS]

- 1. Wash adherent cells twice in the dish or flask with ice-cold PBS and drain off PBS. Wash non-adherent cells in PBS and centrifuge at 800 to 1000 rpm in a table-top centrifuge for 5 minutes to pellet the cells.
- 2. Add ice-cold modified RIPA buffer to cells (1 mL per 10 cells/100 mm dish/150 cm flask; 0.5 mL per 5 x 10 cells/60 mm dish/ 75 cm flask).
- 3. Scrape adherent cells off the dish or flask with a plastic cell scraper. Transfer the cell suspension into a centrifuge tube, and pass 10~20 times through a 21 gauge needle.
- 4. Centrifuge the lysate at 14,000 x g in a pre-cooled centrifuge for 15 minutes. Immediately transfer the supernatant to a fresh centrifuge tube and discardthe pellet.
- 5. To prepare Protein A/G-Agarose, wash the beads twice with PBS and restore to a 50% slurry with PBS. It is recommended to cut the tip off of the pipette when manipulating agarose beads to avoid disruption of the beads.
- 6. Pre-clear the cell lysate by adding 20 μl of Protein A/G-Agarose slurry (50%) per 1 mL of cell lysate and incubating at 4 °C for 10 minutes on a rotator. Pre-clearing the lysate will reduce non-specific binding of proteins to the agarose when it is used later on in the assay.
- 7. Remove the Protein A/G-Agarose by centrifugation at 14,000 x g at 4°C for 5 minutes. Transfer the supernatant to a fresh centrifuge tube.
- 8. Determine the protein concentration of the cell lysate (e.g. if performing a Bradford assay one must dilute the cell lysate at least 1:10 before determining the protein concentration because of the interference of the detergents in the lysis buffer with the Coomassie-based reagent).



- 9. Dilute the cell lysate to approximately 1 μg/μl total cell protein with PBS to reduce the concentration of the detergents in the buffer. A more concentrated cell lysate (i.e., 10 μg/μl) may be necessary to immunoprecipitate a protein, which is found in low levels in a cell model.
- 10. Add the recommended volume of the immunoprecipitating antibody (see antibody datasheet for detailed information) to 500 i.e., 500 g) of cell lysate.
- 11. Gently rotate the cell lysate/antibody mixture for either 2 hours or overnight at 4°C on a rotator.
- 12. Capture the immunocomplex by adding 30-40 μl Protein A/G-Agarose slurry (15-20 μl packed beads) and gently rotating on a rotator for 1 or 2 hour at 4°C.
- 13. Collect the agarose beads by centrifugating for 3 minutes at 1000 rpm. Discard the supernatant and wash the beads 3 times with 800 µl icecold RIPA buffer (more stringent) or PBS (less stringent).
- 14. Resuspend the agarose beads in 30-60 µl 1x SDS loading buffer and mix gently.
- 15. The agarose beads are boiled for 10 minutes at 100°C to dissociate the immunocomplexes from the beads. The beads are collected by centrifugation and SDS-PAGE is performed with the supernatant. Unused samples may be stored at -20°C for later use.

